The effect of mycorrhization with Pisolithus tinctorius on field growth of micropropagated plants of Arbutus unedo L.



Gomes, F.¹; Machado, H.²; Sorzabalbere, I.¹; Moreira, F.¹ & Canhoto, J.M.³ ¹⁻ CERNAS, Escola Superior Agrária de Coimbra, ESAC/IPC, Bencanta P 3040-316 Coimbra, Portugal ²⁻ INRB, Instituto Nacional de Recursos Biológicos, IP/L-INIA, Av. República, Quinta do Marquês 2780-159 Oeiras, Portugal ³⁻ Centre for Functional Ecology, Department of Life Sciences, University of Coimbra, Ap. 3046, 3001-401 Coimbra, Portugal

INTRODUCTION

•Arbutus unedo L. (strawberry tree) grows spontaneously in Mediterranean ecosystems. Fruit production represents the major income for farmers. Adult plants have been selected for fruit production (Fig. 1) and micropropagated (Gomes & Canhoto 2009). Mycorrhizae can improve plant adaptation and tolerance to several types of biotic and abiotic stresses.

•Species of Arbutus (Ericaceae) form arbutoid mycorrhizae with several fungi which are also able to form ectomycorrhizae in association with other plant hosts (Massicotte et al., 1993). In this work the effect of mycorrhization with Pisolithus tinctorius on field growth of micropropagated plants was evaluated

Fig. 1 - Arbutus unedo L. (strawberry tree)

MATERIALS AND METHODS

•Fungal isolates of Pisolithus tinctorius were obtained from sporocarps tissues collected in forest stands of Quercus suber and Eucalyptus globulus.

•Isolations were carried out through the culture of sporocarps tissues on Modified Melin Norkrans (MMN) agar culture medium at 30°C (Jacob et al. 2001). Pure cultures were obtained in MMN liquid medium (glucose 1% w/v) at 30°C. The mycelium was diluted in water (1:20; v/v), and then applied to the substrate (Fig. 2 - d, e, g).

•In vitro cloned plants of A. unedo through shoot proliferation were inoculated in the nursery with two inoculation treatments and compared to control plants (Tab. 1).

•Micropropagated plants (clone C1) were watered with either P. tinctorius mycelium (pure cultures in MMN liquid medium) or dry sporocarps (Tab. 1).

• At the nursery, before plantation roots were checked for the presence of mycorrhizae development.

•A field trial was established to test all the nursery inoculation treatments and fertilized plants: seedlings and 3 micropropagated clones (Fig. 2 - h).

•To study the effect of the inoculation treatments on plant growth, height was evaluated 2 and 4 months in the nursery and 20 months after plantation (Fig. 2 - i).

Table 1 - Inoculation treatments with Pisolithus tinctorius and the control

Type of inoculum tested in vitro clonal plants (C1)				
C1M	Vegetative inoculum (mycelium) from sporocarps from a Quercus suber stand			
C1S	Dry sporocarps from a Eucalyptus globulus stand			
C1C	Control micropropagated plants			

RESULTS

•Two months after inoculation no significant differences were found between mycorrhizal treatments and control plants in the nursery (Tab. 2).

•After 4 months in the nursery best results were achieved with dry sporocarps treatment (C1S) (P<5%; Tab. 2). The control plants showed the lowest height increment value (Δ H; Tab. 2).

•At the nursery, before plantation the plants inoculated (C1M and C1S) showed mycorrhized roots.

•The survival rate in the field trial was about 96.8%, without significant differences between treatments (12 months after plantation).

•Twenty months after field trial establishment both mycorrhizal treatments tested in nursery improved plant growth (Tab. 3), but results were not significantly different.

•Seedlings and clone AL2 showed the lowest height average, even though they have been fertilized.

•Control plants (C1C) showed the lowest average of height increment after 20 months (Table 3; P>0.05)

•Inoculated plants in nursery (C1M and C1S) showed averages of height (H) and height increment (Δ H) higher than the control plants (P>5%; Tab. 3).

CONCLUSIONS

•Addition of fertilizers is a common agricultural practice in field plantations by using a slow release fertilizer. Mycorrhization showed to be a more interesting option since it is cheaper, cleaner and improves the physiological conditions of the plants.

• Further studies are needed for fungal strains selection for their aggressiveness under nursery and field conditions, which is a requirement for a successful implementation of these methods.

•Mycorrhizal synthesis with edible fungi can improve not only the success of plant growth, but can also account for another source of income, for farmers.

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Fig. 2 - Inoculation of micropropagated plants with P. tinctorius in nursery. A. unedo cultures were in vitro multiplied (a). Plantlets were acclimatized (b) and then transferred to containers (c). Following growth of P. tinctorius vegetative inoculum, the mycelium and liquid medium were mixed (d) and diluted with water (e) for application to the acclimatized plants (f). After mycorrhization treatments (g) a field trial was established (h) and plant growth was evaluated (i).

Table 2 – Effect of different inoculation treatments with P. tinctorius on plants' height increment (AH) evaluated 2 and 4 months after inoculation under nursery conditions.

Treatments	Δ H 2 months (cm)	Δ H 4 months (cm)
C1 C - control plants	$10.7{\pm}0.4^{a}$	14.4 ± 0.7^{b}
C1 M - with mycelium in vitro produced	11.0±0.4ª	16.0±0.6 ^b
C1 S - dry sporocarps water mixed	10.8±0.3ª	19.1±0.7ª

In each column values (mean ± SE) followed by different letters are significantly different (P≤0.01).

Table 3 - Effect of the different treatments on plants height and height increment (ΔH) evaluated 20 months after field trial establishment

Mycorrhizal treatments	H (20 months)	$\Delta \mathbf{H} (\mathbf{H}_{20} - \mathbf{H}_{0})$
	mean ± SE (cm)	mean ± SE (cm)
AL1 - selected clone	76.6 ± 7.2^{a}	58.2 ± 7.7^{a}
AL2 - selected clone	68.6 ± 5.8^{a}	55.1 ± 5.8^{a}
AL3 - selected clone	75.3 ± 5.7^{a}	63.2 ± 6.0^{a}
SE - seedlings	68.6 ± 5.3^{a}	57.7 ± 4.8^{a}
C1 C - control plants	76.2 ± 6.7^{a}	54.8 ± 6.9^{a}
C1 M - mycelium in vitro produced	83.0 ± 4.7^{a}	62.0 ± 4.6^{a}
C1 S dry sporospress water mixed	847 ± 51^{a}	608 ± 50^{8}

In each column values (mean±SE) followed by same letters are not significantly different (P≤0.05).

REFERENCES

Gomes, F., Canhoto, J.M., 2009. Micropropagation of strawberry tree (Arbutus unedo L.) from adult plants. In Vitro Cell. Dev. Biol.-Plant Volume 45, 72-87

72-82. Jacob C, Courbot M, Brun A, Steinman HM, Jacquot JP, Botton B, Chalot M (2001) Molecular cloning, characterization and regulation by cadmium of a superoxide dismutase from the extomycorrhizal fungus Paciflus involutus. Eur J Biochem 268:3223-322. Massicotte HB, Melville LH, Molina R, Peterson RL (1993) Structure and histochemistry of mycorrhiza synthesized between Arbutus menziesii (Ericcease) and two basidomycetes Pisolihuus interorius (Pisolihaceae) and Piloderma bicolor (Corciticaee). Mycorrhiza 31-11.
Parlad J, Pera J, Luque J (2004) Evaluation of mycelial inocula of edithe Lactarius species for the production of Pinus pinuster and P. sylvestris mycorrhizal seedings under greenhouse conditions. Mycorrhiza 14:171-176.
Rincón A, Alvarez IF, Joan Pera J (2001) Inoculation of containerized Pinus pinea L. seedlings with seven ectomycorrhizal fungi. Mycorrhiza 11:265-721

. A, Parladé J, Pera J (2005) Effects of ectomycorrhizal inoculation and the type of substrate on mycorrhization, growth and nutrition of tainerized *Pinus pinea* L. seedlings produced in a commercial nursery. Ann For Sci 62:817-822. Rin